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The Importance of Family-genetic Screening: The Phenotype Caused by the p.L3778F Ryanodine Receptor Mutation is Likely Less Severe Than Previously Thought

La importancia del estudio familiar y genético: la mutación p.L3778F en el receptor de la rianodina probablemente no cause un fenotipo tan grave

To the Editor,

We present the case of a family with a history of sudden cardiac death at a young age. The initial diagnosis was catecholaminergic polymorphic ventricular tachycardia (CPVT) caused by the p.L3778F mutation in the cardiac ryanodine receptor gene (*RyR2*). Several years after this initial diagnosis, a comprehensive family genetic study by next generation sequencing (NGS) has now identified a second pathogenic mutation, this time in *KCNQ1* and linked to type 1 long QT syndrome (LQTS1). The severe phenotype

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is likely due to the presence of both mutations and not exclusively to the p.L3778F mutation in *RyR2* as previously reported.¹

The proband (III:5) had a cardiac arrest at the age of 8 years while swimming but recovered fully after resuscitation. His brother had died suddenly, also while swimming, at 10 years of age (no abnormalities were detected on autopsy).

An electrocardiogram (ECG) of the proband showed sinus bradycardia with a QTc of 440 ms, whereas the results of echocardiography, Holter recording, and exercise stress testing were normal. The context of the cardiac arrest suggested LQTS1, and the patient was prescribed beta-blockers. An electrophysiological study was conducted without arrhythmia induction, and the patient was fitted with a subcutaneous Holter monitor. At the age of 10 years, he had an exertion-induced syncopal episode, and ECG revealed self-limiting polymorphic ventricular tachycardia (Figure A).

The patient was referred for further investigation to a center with expertise in cardiac channelopathies. A mutational screen was conducted of the *RyR2*, *KCNQ1*, *KCNH2*, *SCN5A*, *KCNE1*, and



Figure. A: Recording of the polymorphic tachycardia in the proband. B: Family tree./, deceased; –, noncarriers, lacking the L3778F mutation in *RyR2* (blue) or the E449R*14 mutation in *KCNQ1* (red); +, carriers, heterozygous for L3778F in *RyR2* (blue) or E449R*14 in *KCNQ1* (red); circles, women; boxes, men; arrow, proband; blue fill, patient with CPVT (according to the European and US consensus statement)³; black fill, patient clinically affected by LQTS1 and CPVT; red fill, patient with LQTS1; *RyR2*, ryanodine receptor gene; LQTS1, type 1 long QT syndrome; CPVT, catecholaminergic polymorphic ventricular tachycardia.

Table

Characteristics of Carriers of the Mutations in KCNQ1 or RyR2

Sex	Age at diagnosis, y	Syncopal episodes	QTc, ms	Mutation Glu449Arg*14 (KCNQ1)	Mutation Leu3778Phe (RyR2)
F	86	No	476	+	-
М	67	No	446	+	-
М	65	No	461	+	-
F	64	No	474	+	-
F	62	No	467	+	-
М	57	No	483	+	-
F	28	No	446	+	-
М	63	No	432	-	+
М	10	Sí	440	+	+
F	39	No	416	-	+
М	5	No	406	-	+
	Sex F M F F F M F M M F M	Sex Age at diagnosis, y F 86 M 67 M 65 F 64 F 62 M 57 F 28 M 63 M 10 F 39 M 5	SexAge at diagnosis, ySyncopal episodesF86NoM67NoM65NoF64NoF62NoM57NoF28NoM63SíM10SíF39NoM5No	Sex Age at diagnosis, y Syncopal episodes QTc, ms F 86 No 476 M 67 No 446 M 65 No 461 F 64 No 474 F 62 No 467 M 57 No 483 F 28 No 432 M 63 No 432 M 10 Sí 440 F 39 No 416 M 5 No 406	SexAge at diagnosis, ySyncopal episodesQTc, msMutation Glu449Arg*14 (KCNQ1)F86No476+M67No446+M65No461+F64No474+F62No467+M57No483+F28No446+M63No432-M10Sí440+F39No416-M5No406-

F, female; M, male; *RyR2*, ryanodine receptor gene.

KCNE2 genes by denaturing high performance liquid chromatography (DHPLC) and Sanger sequencing, identifying a heterozygous missense mutation in *RyR2* (p.L3778F). Beta-blockers were prescribed, and the patient underwent left sympathetic denervation and was fitted with an implantable cardioverter-defibrillator. The patient's immediate family members were asymptomatic, but the father and sister were identified as carriers of the mutation (Figure B). The mutation had not been described before and the case was included in a study of CPVT patients that highlighted the poor prognosis of male carriers.¹

Several years later, the family relocated and the proband's 5year-old niece (IV:1) was referred for evaluation. She was asymptomatic, but the study confirmed that she carried the family mutation. Interestingly, her 63-year-old grandfather (II:7) and 39-year-old mother (III:6) were both carriers and yet were asymptomatic on ECG, exercise stress testing, and Holter recording (except for an exercise stress test in the grandfather that showed an isolated premature ventricular contraction in 2 morphologies upon exercise initiation, which disappeared with exertion).

Suspicion of a second mutation prompted a new genetic study of the proband by NGS (195 genes). In addition to the p.L3778F mutation in *RyR2*, a second mutation was detected, in *KCNQ1* (g.2610034_2610035insC); this mutation generates an aberrant transcript with a stop codon in amino acid position 463 (p.E449R*14). The mutation had been previously reported in relation to LQTS1,² but was not detected in the original genetic analysis by DHPLC and Sanger sequencing.¹

Further analysis in partnership with other centers confirmed the *KCNQ1* mutation in 7 family members (Figure B). All affected individuals were asymptomatic, yet the mean QTc on basal ECG was 460 ± 15 ms (Table). Given that 25% of LQTS1 patients can have a normal QTc on basal ECG, the carriers of this mutation were diagnosed with LQTS1, according to current diagnostic criteria.³

Beta-blockers were prescribed to carriers of either mutation, and drugs that prolong QT interval were contraindicated for carriers of the *KCNQ1* mutation. At the time of writing, all carriers have remained asymptomatic except for the proband, who is the only carrier of both mutations and had an appropriate implantable cardioverter-defibrillator discharge when aged 19 years.

In summary, the phenotype associated with the p.L3778F *RyR2* mutation in this family is probably not as severe as initially suspected. However, until further studies are conducted, it is premature to discount a pathogenic effect for this mutation, especially given that exercise stress test results can be normal in individuals with CPVT.⁴ The disease severity in the proband, and probably also in his deceased brother, is likely due to the presence

of both mutations. The DHPLC-Sanger method can give false negatives, a limitation overcome with genetic screening by NGS.⁵ The current study highlights the incomplete penetrance and variable expression of LQTS1,^{3,6} the value of genetic testing for detecting asymptomatic carriers,^{3,6} and the importance of intercenter cooperation to ensure a complete family study.

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Unexpected Interrupted Inferior Vena Cava Diagnosed During Failed Transcatheter Left Atrial Appendage Closure

Diagnóstico inesperado de una interrupción de la vena cava inferior durante el fallo de un cierre percutáneo de la orejuela auricular izquierda

To the Editor,

An 88-year-old man with a history of biological aortic valve replacement, pacemaker implantation and chronic atrial fibrillation with thromboembolic protection (dabigatran 110 mg x 2) was admitted for serious intestinal bleeding. Bowel angiodysplasia was diagnosed and the anticoagulation therapy was discontinued. The patient was scheduled to undergo endovascular left atrial appendage (LAA) closure with the Amplatzer Amulet device (St Jude Medical)¹ given that the patient had a score of 4 on both the HAS-BLED and CHA₂DS₂-VASc scales. After sheath insertion in the right femoral vein, a 0.035" conventional J-tip wire repeatedly crossed to the left side of the spine. Contrast media injection confirmed the absence of a right inferior vena cava (IVC) and showed a left IVC combined with a hemiazygos continuation (Figure 1), confirmed by computed tomography angiography (Figure 2). Consequently, the procedure was aborted and finally cancelled. Of note, the transgastric IVC long-axis view by transesophageal echocardiography showed the hepatic veins and strongly suggested an IVC interruption (Figure 1).

All isolated congenital variations of the IVC are a consequence of abnormal embryologic development, affecting approximately



Figure 1. A: Venography showing a left IVC (arrow) and the absence of the right IVC. B: Hemiazygos continuation (2 arrows) and, due to the azygos cross (asterisk), the inferior systemic venous return enters the superior vena cava (arrow head and limited by the red line). C: Transgastric IVC long-axis view by transesophageal echocardiography. Superior vena cava (white star); microbubbles in the left atrium coming from the superior vena cava after injection in the left IVC (red star). D: IVC (white star), hepatic veins (white arrows); IVC interruption (red arrows). IVC, inferior vena cava.