

Editorial

Associating Chagasic Cardiomyopathy With Abnormal Diastolic Calcium Handling

Asociación de la miocardiopatía chagásica con el comportamiento anormal del calcio diastólico

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Calcium is a central player in the regulation of cardiac contraction and rhythm, and predominant cardiac pathologies have been associated directly or indirectly with changes in intracellular calcium handling.^{1–4} A feature common to many diseases involving either rhythm disturbances or impaired cardiac function is that they are linked to genetic mutations^{2,5} or abnormal phosphorylation of one or several calcium handling proteins^{4–7} that alter their activity. In patients with heart failure, multiple alterations in calcium handling have been reported that may account for the impaired contraction and relaxation^{3,8} as well as the propensity of these patients to present ventricular arrhythmia.⁴ It is therefore natural to speculate that the tropical disease known as Chagas disease caused by the hemoflagellate *Trypanosoma cruzi*, which eventually induces dilated cardiomyopathy, systolic and diastolic dysfunction, arrhythmias, and sudden cardiac death,⁹ also induces changes in the calcium metabolism in cardiac myocytes from chagasic patients.

So far, studies on potential effects of chagasic infection on calcium handling have been carried out in noncardiac preparations from animal models. Therefore, to address this important issue in humans, in the article published in *Revista Española de Cardiología*, López et al.¹⁰ have undertaken the difficult task of measuring the diastolic calcium concentration in ventricular myocytes from patients infected with *Trypanosoma cruzi*, at different stages of the progression of this disease.

Since the development of fast responding fluorescent calcium indicators with high quantum efficiency,¹¹ these compounds have been widely used to measure and visualize changes in cytosolic calcium levels in isolated cardiomyocytes and multicellular preparations. However, it is commonly overlooked that the chemical properties of most fluorescent calcium indicators make it difficult to use them for quantification of the cytosolic calcium concentration. Thus, the fluorescent calcium indicators represent exogenous calcium buffers with calcium affinities that typically fall between the diastolic and systolic calcium levels. This is bound to affect the cellular calcium homeostasis, and the problem is aggravated by the fact that common calcium indicators such as

fura-2, indo-1 and fluo-3 are known to bind extensively to cellular proteins, which increases their effective calcium buffering capacity dramatically.¹² Moreover, the binding of these compounds to cellular proteins substantially alters their calcium affinity,^{12,13} further complicating their use for quantitative purposes. In contrast to this, calcium selective electrodes have the virtues that make them ideal for measurements of static calcium concentrations; ie, they have a wide dynamic range, they do not interfere with the cytosolic calcium level, and their calibration is fairly uncomplicated. The drawbacks that have pushed the calcium electrodes into the shadow of the fluorescent calcium dyes is their slow response time and the necessity of impaling the myocyte under study, making it a demanding experimental technique with a limited success rate.¹⁴ However, for quantitative purposes in quiescent preparations calcium selective electrodes remain superior to the fluorescent dyes. A further advantage of this technique is that it allows simultaneous determination of the resting membrane potential.

Using calcium-selective microelectrodes, López et al.¹⁰ measured a resting membrane potential of -83 ± 2 mV in 30 ventricular myocytes from 4 control patients, a value that compares well to determinations in multicellular human ventricular preparations.¹⁵ The diastolic calcium level in these same 30 myocytes was 111 ± 4 nM. This value also agrees with estimations of the diastolic calcium level in human ventricular myocytes using the fluorescent calcium indicator fluo-3,³ affirming that calcium-selective microelectrodes is a useful technique to simultaneously record the resting membrane potential and diastolic calcium levels in human ventricular myocytes. The calcium-selective microelectrodes were therefore used to impale human ventricular myocytes from patients at different stages of the progression of Chagas disease, and the results provide evidence that progression of the disease is associated with a parallel increase in the diastolic calcium concentration. Although not directly addressed in the study of López et al.,¹⁰ this is likely to cause calcium overloading of the sarcoplasmic reticulum and it may contribute to the propensity of chagasic patients to suffer cardiac arrhythmia. The higher diastolic calcium level in ventricular myocytes from these patients is also expected to reduce the dynamic range for modulation of the calcium transient amplitude and this likely contributes to promote diastolic and systolic dysfunction. Furthermore, the concurrent loss of the resting membrane potential, which reduces excitability, may exacerbate the effects of diastolic calcium overload in patients with Chagas disease.

To determine whether the abnormal diastolic calcium handling in chagasic patients is associated with changes in inositol

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triphosphate (IP₃) signaling, López et al.¹⁰ used pharmacological manipulation of cellular IP₃ production and IP₃ receptor activation. Their results show that blockade of IP₃ production (with the β -phospholipase C inhibitor U73112) selectively reduced the diastolic calcium concentration in chagasic patients without changing the resting membrane potential, while stimulation of IP₃ production with phenylephrine had the opposite effect on diastolic calcium. IP₃ receptor blockade (with 2-APB) reduced the elevated diastolic [Ca²⁺] in functional class I and II chagasic patients towards concentrations observed in control patients, and reversed the stimulatory effect of phenylephrine, suggesting that there is an upregulation of IP₃ receptor-mediated signaling in ventricular myocytes from patients with Chagas disease.

Together, these findings suggest that upregulation of IP₃ receptor-mediated signaling and abnormal diastolic calcium handling are novel mechanisms that may contribute to promote diastolic dysfunction in patients with Chagas disease. The study also opens new lines of research needed to 1) address the molecular mechanism underlying the progressive loss of the resting membrane potential in these patients, 2) determine if abnormal diastolic calcium handling is associated with the loss of the membrane potential, and 3) establish how the observed elevation of the diastolic [Ca²⁺] is linked to upregulation of IP₃ receptor activation. Another finding of the study by López et al. that deserves further investigation is the dramatic elevation of the diastolic [Ca²⁺] observed in class III patients (922 ± 33 nM) without any apparent cell contracture. The authors discuss different explanations for this observation but a tempting hypothesis is that there is a parallel decrease in the calcium sensitivity of the myofilaments during progression of Chagas disease.

The importance of the findings reported by López et al.¹⁰ is unquestionable, and as mentioned above their results provide a foundation for future research on the mechanisms underlying abnormal diastolic calcium handling in chagasic diseased cardiomyocytes. However, I find it of particular merit that the authors have taken the effort to use the best-suited, albeit experimentally most demanding technique, to measure the diastolic [Ca²⁺] in human ventricular myocytes – a highly relevant but technically difficult experimental preparation.

CONFLICTS OF INTEREST

None declared.

REFERENCES

1. Hove-Madsen L, Llach A, Bayes-Genis A, Roura S, Rodriguez Font E, Arís A, et al. Atrial fibrillation is associated with increased spontaneous calcium release from the sarcoplasmic reticulum in human atrial myocytes. *Circulation*. 2004;110:1358–63.
2. Jiang D, Xiao B, Yang D, Wang R, Choi P, Zhang L, et al. Ryr2 mutations linked to ventricular tachycardia and sudden death reduce the threshold for store-overload-induced Ca²⁺ release (SOICR). *Proc Natl Acad Sci USA*. 2004;101:13062–7.
3. Piacentino V 3rd, Weber CR, Chen X, Weisser-Thomas J, Margulies KB, Bers DM, et al. Cellular basis of abnormal calcium transients of failing human ventricular myocytes. *Circ Res*. 2003;92:651–8.
4. Reiken S, Wehrens XH, Vest JA, Barbone A, Klotz S, Mancini D, et al. Beta-blockers restore calcium release channel function and improve cardiac muscle performance in human heart failure. *Circulation*. 2003;107:2459–66.
5. Erxleben C, Liao Y, Gentile S, Chin D, Gomez-Alegria C, Mori Y, et al. Cyclosporin and timothy syndrome increase mode 2 gating of cav1.2 calcium channels through aberrant phosphorylation of S6 helices. *Proc Natl Acad Sci USA*. 2006;103:3932–7.
6. Llach A, Molina CE, Prat-Vidal C, Fernandes J, Casado V, Ciruela F, et al. Abnormal calcium handling in atrial fibrillation is linked to up-regulation of adenosine a2a receptors. *Eur Heart J*. 2011 Mar;32:721–9.
7. Sossalla S, Fluschnik N, Schotola H, Ort KR, Neef S, Schulte T, et al. Inhibition of elevated ca2+/calmodulin-dependent protein kinase ii improves contractility in human failing myocardium. *Circ Res*. 2010;107:1150–61.
8. Kubo H, Margulies KB, Piacentino V 3rd, Gaughan JP, Houser SR. Patients with end-stage congestive heart failure treated with beta-adrenergic receptor antagonists have improved ventricular myocyte calcium regulatory protein abundance. *Circulation*. 2001;104:1012–8.
9. Rassi Jr A, Rassi A, Little WC. Chagas' heart disease. *Clin Cardiol*. 2000;23:883–9.
10. López JR, Espinosa R, Landazaru P, Linares N, Allen P, Mijares A. Disfunción de la [Ca²⁺] diastólica en cardiomiocitos aislados de pacientes chagásicos. *Rev Esp Cardiol*. 2011;64:456–62.
11. Grynkiewicz G, Poenie M, Tsien RY. A new generation of Ca²⁺ indicators with greatly improved fluorescence properties. *J Biol Chem*. 1985;260:3440–50.
12. Hove-Madsen L, Bers DM. Indo-1 binding to protein in permeabilized ventricular myocytes alters its spectral and ca binding properties. *Biophys J*. 1992;63:89–97.
13. Harkins AB, Kurebayashi N, Baylor SM. Resting myoplasmic free calcium in frog skeletal muscle fibers estimated with fluo-3. *Biophys J*. 1993;65:865–81.
14. Hove-Madsen L, Baudet S, Bers DM. Making and using calcium-selective mini- and microelectrodes. *Methods Cell Biol*. 2010;99:67–89.
15. Jakob H, Nawrath H, Rupp J. Adrenoceptor-mediated changes of action potential and force of contraction in human isolated ventricular heart muscle. *Br J Pharmacol*. 1988;94:584–90.