REVIEW ARTICLE

Rho/Rho Kinase Signal Transduction Pathway in Cardiovascular Disease and Cardiovascular Remodeling

Jorge Jalil,^a Sergio Lavandero,^{b,c} Mario Chiong,^{b,c} and María Paz Ocaranza^a

^aDepartamento de Enfermedades Cardiovasculares, Laboratorio de Cardiología Molecular, Hospital Clínico, Pontificia Universidad Católica de Chile, Chile.

^bDepartamento de Bioquímica y Biología Molecular, Facultad de Ciencias Químicas y Farmacéuticas,

Universidad de Chile, Chile.

°Centro FONDAP Estudios Moleculares de la Célula, Universidad de Chile, Chile.

The small guanosine triphosphatase Rho and its target, Rho kinase, play important roles in both blood pressure regulation and vascular smooth muscle contraction. Rho is activated by agonists of receptors coupled to cell membrane G protein, such as angiotensin II and phenylephrine. Once Rho is activated, it translocates to the cell membrane where it, in turn, activates Rho kinase. Activated Rho kinase phosphorylates myosin light chain phosphatase, which is then inhibited. This sequence stimulates vascular smooth muscle contraction, stress fiber formation, and cell migration. In this way, Rho and Rho kinase activation have important effects on several cardiovascular diseases. Currently available substances that specifically inhibit this signaling pathway could offer clinical benefits in several cardiovascular, as well as non-cardiovascular, diseases, such as arterial hypertension, pulmonary hypertension, cerebral or coronary spasm, post-angioplasty restenosis, and erectile dysfunction.

Key words: Rho. Rho kinase. Small G proteins. Arterial hypertension. Cardiovascular remodeling.

La vía de señalización Rho/Rho-cinasa en la enfermedad y el remodelado cardiovascular

La guanosina trifosfatasa pequeña Rho y su blanco Rho-cinasa desempeñan un papel importante en la regulación de la presión arterial y en la contracción de la musculatura lisa vascular.

Diversos agonistas de los receptores acoplados a la proteína G de la membrana celular como, por ejemplo, la angiotensina II y la fenilefrina, activan Rho, que se transloca así hacia la membrana plasmática donde activa Rho-cinasa.

Al activarse Rho-cinasa por RhoA se fosforila la fosfatasa de la cadena ligera de miosina, con lo cual esta fosfatasa es inhibida, y se favorece de esta manera la contracción de las células musculares lisas vasculares, la formación de fibras de estrés y la migración celular. Así, la activación de Rho y de Rho-cinasa tiene efectos importantes en diversas enfermedades cardiovasculares.

Al disponer en la actualidad de inhibidores de esta vía de señalización, diversas enfermedades cardiovasculares (y también otras no cardiovasculares) se podrían beneficiar clínicamente, entre ellas la hipertensión arterial, la hipertensión pulmonar, el espasmo cerebral y coronario, la reestenosis postangioplastia y la disfunción eréctil.

Palabras clave: *Rho. Rho-cinasa. Proteínas G pequeñas. Hipertensión arterial. Remodelado cardiovascular.*

INTRODUCTION

Arterial hypertension is a cardiovascular disorder characterized by altered vascular tone and increased

Correspondence: Dr. J. Jalil.

Departamento de Enfermedades Cardiovasculares. Hospital Clínico. Pontificia Universidad Católica de Chile. Lira, 85, piso 2. Santiago de Chile. Chile.

E-mail: jjalil@med.puc.cl

vascular contractility. It is accompanied by proliferation and migration of vascular smooth muscle cells and differing degrees of inflammation of the arterial wall, processes that together constitute vascular remodeling. Proliferation, migration of vascular smooth muscle cells from the media to the intima, and inflammation of the arterial wall participate in a number of vascular disorders, including for example, the progression of atherosclerosis, restenosis, rejection of bypass grafts, and renal failure. In addition to cardiac remodeling, arterial hypertension is accompanied by hypertrophy of cardiac myocytes, hyper-

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Figure 1. Activation-inactivation cycle of the small GTPases. External factors activate quanine nucleotide exchange factor (GEF). This factor displaces GDP dissociation inhibitor (GDI), releasing the isoprenylated residue that anchors the small GTPase to the plasma membrane, and catalyzes the exchange of GDP for GTP. GTP binding induces a conformational change in the small GTPase that activates it and allows it to bind effectors. GTPase activating protein (GAP) stimulates the intrinsic GTPase activity of the small GTPase. GTP hydrolysis returns the small GTPase to its inactive state bound to GDP and GDL

plasia of fibroblasts, and alterations of the extracellular matrix.

Of the main complications of arterial hypertension, those associated with the development of atherosclerosis are the least affected by current antihypertensive treatment. A metaanalysis of the effects of antihypertensive treatment revealed a 48% reduction in the incidence of cerebrovascular accidents compared with only a 16% reduction in the incidence of heart disease,¹ figures that are markedly lower than would have been predicted from available epidemiological studies. This finding can be interpreted in a number of ways. One of these is that development of atherosclerosis and the associated complications are not effectively prevented by current antihypertensive treatment. These findings are consistent with the lack of evidence to support lowering of arterial blood pressure as an effective mean of reducing atherosclerosis or proliferation of vascular smooth muscle cells, and provide a strong incentive to investigate other mechanisms of vascular damage and remodeling with greater potential for treatment and prevention.

It has recently been shown that activation of the Rho/Rho kinase signal transduction pathway is one of the principal mechanisms of vasoconstriction in arterial hypertension and that this pathway has novel therapeutic potential.²

Studies have suggested that the small guanosine triphosphatase (GTPase) Rho and its target Rho kinase play a crucial role in the regulation of arterial blood pressure.³ In vitro studies have demonstrated that the activated form of Rho kinase regulates vascular

smooth muscle contraction via phosphorylation of the myosin light chain, sensitization of contractile proteins to Ca⁺⁺, and the formation of stress fibers. Furthermore, Rho kinase may also be regulated by various components of the cytoskeleton that play an essential role in the mechanotransduction of flow and pressure in the blood vessels. In addition to a role in the pathogenesis of atherosclerosis, Rho kinase participates in the organization of the actin cytoskeleton, in the processes of cell adhesion and motility, in cytokinesis, and in gene expression.⁴ Prolonged treatment with fasudil (a specific inhibitor of Rho kinase) has been shown to reduce the development of coronary vascular lesions such as medial thickening and perivascular fibrosis in spontaneously hypertensive rats.⁵

THE RHO/RHO KINASE SIGNAL TRANSDUCTION PATHWAY

The small GTPase superfamily contains more than 100 structurally related proteins that undergo conformational changes and alter their subcellular localization in a manner that is dependent on guanine nucleotides. Small GTPases are active when bound to GTP and inactive when bound to GDP. In their activated state, they bind effectors that, in turn, regulate a large number of biological processes. They are controlled by various classes of regulatory proteins.⁶ Figure 1 shows the cycle of activation and inactivation of the small GTPases. With few exceptions, their activation is mediated by guanine nucleotide exchange factors (GEFs). These factors displace the GDP dissocia-

Figure 2. Regulation of cell responses through activation of small GTPases (Ras, RhoA, RhoB, and Rac1). Angiotensin II (Ang II) stimulates Ras RhoA, RhoB, and Rac1 through a process dependent upon the activation of a G-protein coupled receptor at the plasma membrane. The activation of Ras by Gq/11 is probably mediated by protein kinase C (PKC). Ras binds 3 well-defined effectors: phosphatidylinositol 3-kinase (PI3K), RalGDS, and Raf, the latter being one of the first components in the ERK protein kinase cascade. In turn, Raf and RalGDS regulate gene expression associated with hypertrophy, PI3K modulates global gene expression, and the ERK cascade regulates cell survival. RhoA modulates the cytoskeleton and hypertrophy-associated gene expression through the action of Rho kinase. Rac1 activates NAD(P)H oxidase, which controls global gene expression via the generation of the superoxide anion. Rac1, either directly or through NAD(P)H oxidase, also regulates cell



architecture by modulating the actin-myosin cytoskeleton. RhoB has opposing actions to RhoA and inhibits RhoA-mediated gene expression.

tion inhibitor (GDI) and unmask the isoprene group that anchors the small GTPase to the plasma membrane, as well as catalyzing the exchange of GDP for GTP. GTP binding induces a conformational change in the small GTPase that activates it and allows it to bind effectors. GTPase activating proteins (GAP) stimulate the intrinsic hydrolysis of GTP and lead to the rapid conversion of small GTPases to their inactive state bound to GDP and GDI.

Based on structural and functional relationships, the small GTPase superfamily is subdivided into 5 families: Ras, Rho, Rab, Arf, and Ran (Figure 2).⁷

Members of the Ras family include the 3 classical isoforms H-Ras, K-Ras, and N-Ras, and are primarily involved in the regulation of cell proliferation and differentiation. The classical isoforms of Ras also regulate apoptosis. As shown in Figure 2, all of these effects are likely to be regulated by the extracellular regulated kinases (ERK), a subfamily of the mitogenactivated protein kinases (MAPK), or phosphatidylinositol-3-kinase (PI3K).⁸

Members of the Rho family (Rap, Ral, and the Rho subfamily that includes RhoA, RhoB, Rac1, and Cdc42) are unique in controlling the formation of the different conformational forms of the actin cytoskeleton.^{4,8,9} In addition, Rho proteins regulate a large number of other processes, including gene transcription and lipid metabolism. RhoB has opposing actions to RhoA and inhibits RhoA-mediated gene expression. The effects of Rho A on the cellular architecture are mediated by Rho-dependent serine/threonine kinases that fall into 2 subgroups: the related protein kinases protein kinase C (PKC) and protein kinase N, and the Rho kinases.⁸ The latter group, with a mass of approximately 160 kDa, includes ROKa (or ROCK2) and ROKβ (or ROCK 1). ROKα can stimulate LIM-kinase, an enzyme that phosphorylates and inactivates cofilin.¹⁰ Given that cofilin stimulates actin depolymerization,¹¹ the net effect of RhoA through this pathway is to stimulate the formation of actin fibers.

When Rho kinase is activated by RhoA it phosphorylates myosin light chain phosphatase (MLCP; Figure 3),⁴ thereby inhibiting it and favoring the contraction of vascular smooth muscle cells, the formation of stress fibers, and cell migration. The availability of Y-27632, a selective inhibitor of Rho kinase¹² (Figure 4), as well as dominant negative forms of the enzyme, have been useful in elucidating the roles of this pathway.



Figure 3. Regulation of vascular smooth muscle cell contraction by Rho kinase. Regulators of muscle cell contraction activate 3 independent processes. The classical mechanism involves activation of phospholipase C (PLC), production of inositol-1,4,5-triphosphate (IP3), increased levels of intracellular calcium as a result of calcium channel activation, and activation of myosin light chain kinase (MLCK). Phosphorylation of myosin induces and maintains permanent contraction of actinmyosin fibers. The second mechanism functions through PLC-mediated activation of protein kinase C (PKC), which in turn phosphorylates and activates CPI-17. CPI-17 inhibits myosin light chain phosphatase (MLCP), maintaining myosin in a phosphorylated and contracted form. The final mechanism involves activation of guanine exchange factor (GEF), RhoA, and Rho kinase. Rho kinase phosphorylates MLCP and inactivates it.

The processes regulated by the different families of small GTPases are extremely diverse. In fact, there is almost no area of cell biology or biomedical science in which a role for these proteins has not been identified. Furthermore, there are a large number of interactions between members of the Ras and Rho families. The signal transduction pathways regulated by members of the Rho family play in important role in a number of pathologic conditions, including cancer, inflammation, bacterial infection, and arterial hypertension.¹³⁻¹⁵ Rac also controls the generation of reactive oxygen species (ROS), both in leukocytes and nonhematopoietic cells. In the latter cell type, Rac stimulates the production of ROS by activating a protein complex similar to NADPH-oxidase, which has mainly been characterized in neutrophils.¹⁶

These GTPases act as molecular controls in a wide variety of cell processes, among which the best characterized is the regulation of the actin cytoskeleton. Some actions of G-protein-coupled receptors are mediated by small GTPases like Rho. It has been demonstrated that agonists of G-protein-coupled receptors such as angiotensin II, lysophosphatidic acid, carbachol, and phenylephrine increase the concentration of Rho at the membrane and reduce its levels in the cytosol, indicating translocation and activation of Rho.¹⁷

The Rho signal transduction pathway participates in various pathophysiologic processes in the cardiovascu-

lar system.¹⁸⁻²¹ The majority of these processes are directly or indirectly associated with arterial hypertension and its associated complications (Table 1).

INTERACTION BETWEEN ANGIOTENS IN II AND RHO IN THE CARDIOVASCULAR SYSTEM

Vascular remodeling occurs during normal development and participates in various physiologic processes. However, structural changes to the vasculature can be pathologic as well as adaptive and can lead to the development of arterial hypertension, atherosclerosis, and diseased venous bypass grafts. Angiotensin II contributes to vascular remodeling through the activation of signaling cascades that promote vasoconstriction, cell proliferation, and inflammation. The cytoskeleton also participates in adaptive structural responses of the vasculature. The cytoskeleton mediates vasoactive responses and transduces both mechanical stimuli and pharmacologic signals. Some of the cytoskeletal changes that occur in vascular remodeling, specifically in vascular smooth muscle cells, are induced by angiotensin II. In fact, Rho has been directly linked to pathologic vascular remodeling through a series of lines of evidence that demonstrate that angiotensin II activates the Rho/Rho kinase pathway and regulates the cytoskeleton.²² Figure 2 summarizes the angiotensin II-mediated regulation of

TABLE 1. Cardiovascular Processes Involving Rho and Rho Kinase

 Vascular sensitization to calcium through phosphorylation of the myosin light chain, causing arterial vasoconstriction, increased vascular resistance, and arterial hypertension^{3,58,59}
Activation of ERK protein kinases and arterial contraction induced by angiotensin II in resistance vessels^{18,58,59}
Contribution to cerebral arterial tone in chronic arterial hypertension^{19,58}
Mediation of the effects of thrombin on the proliferation and migration of vascular smooth muscle cells¹⁷
Expression of plasminogen activator-1 mRNA, and perivascular and left ventricular myocardial fibrosis in arterial hypertension

induced by chronic administration of angiotensin II²⁰ Early inflammation of the coronary arterial wall and late coronary remodeling in experimental arterial hypertension produced by chronic inhibition of endothelial nitric oxide synthase²¹

cell responses dependent on the activation of GTPases (Ras, RhoA, RhoB, and Rac1).

Cardiac Myocyte Hypertrophy

Angiotensin II evokes a variety of hypertrophic responses in cardiac myocytes, including activation of various protein kinases, reexpression of fetal genes, and increased protein synthesis. In cultured rat cardiac myocytes, angiotensin II activates RhoA, leading to the formation of premyofibrils and the induction of genes associated with the hypertrophic response in a process that appears to differ from that which occurs in striated muscle.23 In these same cells, angiotensin II is a potent activator of ERK proteins. This activation is blocked by the administration of an antagonist of the type 1 angiotensin II receptor, an effect which is prevented by pretreatment of the cardiac myocytes with exoenzyme C3, a potent inhibitor of Rho. These findings suggest that Rho participates in the angiotensin II-mediated activation of ERK proteins in cardiac myocytes.^{24,25} In addition, it has been shown that the activity of the skeletal α -actin and c-fos promoters is upregulated by angiotensin II, and that this effect is partially inhibited by pretreatment with exoenzyme C3. Angiotensin II was found to increase incorporation of tritiated phenylalanine in cardiac myocytes, an effect that was also suppressed by pretreatment with exoenzyme C3. These observations indicate that the Rho family of small GTPases regulates the hypertrophic response of cardiac myocytes to angiotensin II.⁷

Hypertrophy and Hyperplasia of Vascular Smooth Muscle Cells

Rho and Rho kinase, as well as c-fos, play a key role in the hypertrophic changes induced in vascular

smooth muscle cells by angiotensin II.²⁶ In this cell type, Y-27632 abolishes both mRNA and protein expression of c-fos in response to angiotensin II.²⁶ Mechanotransduction in vascular smooth muscle cells depends on the presence of intact actin filaments. Furthermore, Rho is activated by stretching, and the Rho/Rho kinase pathway mediates stretch-induced ERK activation and vascular hyperplasia.²⁷

Angiotensin II, Vascular Inflammation, and RhoA

It has been postulated that Rho kinase could participate in atherogenesis, based on studies of the role of this pathway in vascular inflammation. Monocyte chemoattractant protein-1 (MCP-1) is a chemokine that regulates monocyte recruitment and participates in atherogenesis. Rho kinase regulates the angiotensin-induced production of MCP-1, independently of ERK activation. In vascular smooth muscle cells, this MCP-1 production is blocked by botulinum exotoxin C3.²⁸ Overexpression of a dominant-negative Rho kinase or the use of Y-27632 significantly inhibit the angiotensin II-dependent expression of MCP-1.²⁸

Bradykinins and the Rho Signal Transduction Pathway

Few studies have assessed the possible interaction of bradykinins with the Rho signaling pathway, and even fewer in relation to cardiovascular remodeling. It has been shown in A549 epithelial cells that bradykinin stimulates the activation of NF κ B and the synthesis of interleukin-1 β , and that RhoA is both necessary and sufficient to mediate this effect.²⁹

RHO SIGNALING AND ARTERIAL HYPERTENSION

The role of Rho signaling in arterial hypertension was first recognized in 1997.³ In that study, a specific inhibitor of Rho kinase was observed to reduce arterial blood pressure in 3 experimental models of arterial hypertension. However, the relationship between Rho signaling and pathologic cardiovascular remodeling, or with the vasoactive molecules angiotensin II and the bradykinins, was not studied.

Increases in the expression and activity of RhoA are associated with an increase in DNA synthesis and a reduction in the expression of the cell cycle protein p27/Kip1 in the aorta and tail artery of spontaneously hypertensive and L-NAME-treated rats.³⁰ In the aorta, nitric oxide inhibits the activity of Rho kinase and the sensitization of the tissue to calcium, leading to the hypothesis that nitric oxide-mediated vasodilation occurs through inhibition of the vasoconstrictor activity of Rho kinase.³¹ Nitric oxide induces vasodilation through cyclic GMP. A recent study demonstrated that protein kinase G phosphorylates RhoA and inhibits its activity.³² In addition, sodium nitroprusside reverses the RhoA translocation induced by phenylephrine, indicating inhibition of RhoA activity. Figure 3 summarizes the mechanisms through which RhoA has been observed to cause vasoconstriction in arterial hypertension.

Few studies have addressed the role played by this signal transduction pathway in arterial hypertension in humans. In a recent study in hypertensive patients, fasudil, a specific inhibitor of Rho kinase used in the treatment of cerebrovascular spasm in subarachnoid hemorrhage, was seen to induce a stronger vasodilatory response in the forearm of hypertensive than control subjects, while the response to sodium nitroprusside was similar in both patient groups. This result constitutes the first evidence that the Rho/Rho kinase pathway participates in the pathogenesis of increased systemic vascular resistance in hypertensive patients.³²

THE RELATIONSHIP BETWEEN RHO SIGNALING AND ATHEROGENESIS

The Role of Rho in the Endothelium

There are various interactions between Rho signaling and the structure and function of the endothelium in both normal and pathologic conditions. These interactions participate in the development of arterial hypertension and are associated with the progression of atherosclerosis in various arteries.

The integrity of the endothelium depends on intercellular adhesions mediated by vascular-endothelial (VE) cadherin that, in turn, depend on connections to the actin cytoskeleton. Rho controls cytoskeletal dynamics and cadherin function in epithelial and endothelial cells. The function of these cadherins is also regulated by ROS, which control the pathophysiologic changes associated with inflammation and endothelial damage, and with migration of endothelial cells and angiogenesis.33 Endothelial dysfunction is characterized by an altered endothelial response that favors the processes of vasoconstriction, cell adhesion, and coagulation. The statins, inhibitors of HMG-CoA reductase, are useful for reversing endothelial dysfunction, an effect that appears to be independent of the reduction of serum cholesterol concentration. Rho-dependent activation of preproendothelin-1 expression is inhibited by statins. Thus, control of vascular tone and proliferation mediated by endothelin-1 is regulated at multiple levels, and Rho signaling may play a significant role.34

Rho modulates the permeability of endothelial monolayers and regulates actin filament assembly,

contractile activity due to myosin, and the distribution of the calcium dependent cell adhesion molecule VE cadherin. Rho and the contractile processes mediated by RhoA do not participate in the increased permeability induced by bradykinin and platelet activating factor in intact microvessels.³⁵ The regulation of the endothelial cell barrier is absolutely dependent on components of the cytoskeleton. Factors that cause edema induce contraction of this barrier through actomyosin, associated with phosphorylation of myosin light chain and reorganization of filaments. Alteration of this structural assembly increases dysfunction of the endothelial barrier by activating specific signaling pathways that "talk" to networks of microfilaments and increase the Rhomediated contractility of the endothelial barrier.36 The migration of endothelial cells induced by vascular endothelial growth factor (VEGF) is a critical step in angiogenesis, and requires the participation of Rac, another small GTPase.³⁷

THE RELATIONSHIP BETWEEN RHO SIGNALING AND VASCULAR INFLAMMATION, ADHESION MOLECULES, AND PROCOAGULATION IN ARTERIAL HYPERTENSION

Inflammation is crucial to the pathogenesis of atherogenesis and plays an important role in the vascular complications associated with arterial hypertension. Transmigration of monocytes into the subendothelial space, via integrin activation and chemotaxis, is the initial step in atherosclerotic plaque formation and inflammation. Integrins are activated by ERK, while p38-MAPK and Rho control chemotaxis mediated by MCP-1. Rho and Rho kinase are upstream of p38-MAPK in MCP-1 signaling. Thus, ERK and p38-MAPK regulate distinct signal cascades that lead to integrin activation and chemotaxis induced by MCP-1.38 In addition to regulating fibrinolytic activity, plasminogen activator inhibitor type 1 (PAI-1) also plays a role in the pathogenesis of atherosclerosis and arterial hypertension. Angiotensin II, acting through the type 1 angiotensin II receptor, upregulates protein and mRNA expression of PAI-1 in vascular smooth muscle cells.39 Overexpression of a dominant-negative Rho kinase or the use of Y-27632 completely blocks the induction of PAI-1 expression by angiotensin II, without affecting ERK activation. These observations indicate that activation of the ERK and Rho kinase pathways is necessary for the induction of PAI-1 expression by angiotensin II. Rho kinase could be a novel target through which to inhibit the effects of angiotensin II in the treatment of arterial hypertension and atherosclerosis.

In rat fibroblasts, inhibition of PKC or RhoA selectively inhibits induction of c-fos by transforming growth factor $\beta 1$ (TGF- $\beta 1$), indicating possible roles for both PKC and RhoA in the induction of c-fos by TGF- $\beta 1$.⁴⁰

Rho participates in signaling through chemoattractant receptors that initiates rapid adhesion in leukocytes.⁴¹ The interactions of endothelial ICAM-1 and VCAM-1 with their ligands are involved in the differential regulation of distinct steps in diapedesis by modulating the balance of active and inactive forms of small GTPases.⁴² In endothelial cells of the aorta, the statin cerivastatin blocks lipopolysaccharide-induced expression of ICAM-1 mRNA. Cotreatment with geranylgeranyl pyrophosphate reverses the effect of the statin, indicating that this effect is caused by inhibition of Rho activity.⁴³

Rho mediates the assembly of focal adhesions through integrins and actin stress fibers. Genetic inhibition of RhoA in human endothelial cells has demonstrated a requirement for Rho in the assembly of stable adhesions with monocytes through clustering of the receptors E-selectin, ICAM-1, and VCAM-1, which recognize ligands present on monocytes.⁴⁴

The endothelium of the cerebral blood vessels, which constitutes the blood-brain barrier, controls the adhesion and migration of lymphocytes into the brain. The cell adhesion molecule ICAM-1 participates in the extravasation, morphological changes, and gene regulation associated with the migration of lymphocytes across the blood-brain barrier through activation of Rho and phosphorylation of the endothelial actin cytoskeleton.^{45,46} In the brain, the signaling pathway mediated by ICAM-1 in the endothelial cells is a central element in facilitating lymphocyte migration. Rho proteins, which require posttranslational prenylation to be functionally active, are essential components of this signaling cascade. It has been suggested that the signal transduction systems of cerebral endothelial cells, particularly Rho proteins, could be attractive pharmacological targets through which to recruit leukocytes to the central nervous system.47

It has been observed in experiments with human T lymphocytes that Rho and Rho kinase regulate the tyrosine kinase PYK2 by controlling F-actin-mediated control of the integrin lymphocyte function-associated antigen-1, highlighting a novel form of modulating the activity of this cytoplasmic tyrosine kinase. In addition, Rho regulates the chemokine-induced polarization and migration of lymphocytes, in which cdc42 plays a key role.⁴⁸ Antithrombin directly inhibits chemokine-induced migration of monocytes and lymphocytes via the effects of its heparin binding site by activating PKC and Rho.⁴⁹

Disequilibrium between nitric oxide and endothelin-1 plays an important role in cardiovascular diseases. Thrombin exerts powerful effects on endothelial function. A study has addressed the molecular mechanisms through which thrombin regulates the expression of endothelial nitric oxide synthase (eNOS) and endothelin converting enzyme-1 (ECE-1) in human endothelial cells. Incubation of human umbilical cord endothelial cells with thrombin caused a marked downregulation of eNOS expression and increased the levels of ECE-1 protein in a concentration dependent manner. Thrombin also downregulated and upregulated mRNA expression of eNOS and ECE-1, respectively. The effects of thrombin on eNOS and ECE-1 were found to be mediated by the Rho and ERK pathways, respectively.⁵⁰

NADH/NAD(P)H OXIDASES, REACTIVE OXYGEN SPECIES, AND RHO

The NADH/NAD(P)H oxidase pathway is one of the main producers of ROS in the vascular system. Superoxide (O_2^-) anions are generated that can react with nitric oxide to form the peroxynitrite anion, which lacks the relaxant activity of nitric oxide on the smooth muscle. Angiotensin II induces activation of NADH/NAD(P)H oxidases in vascular smooth muscle cells, leading to increased production of O_2^- and inactivation of nitric oxide. In phagocytes, the NAD(P)H oxidase system that generates O_2^- is regulated by small G proteins related to Rac.⁵¹

In the vascular smooth muscle cells of the rat aorta, inhibition of G α i with pertussis toxin and inhibition of NADH/NAD(P)H oxidase reduces stimulation of p21-activated kinase (PAK) induced by lysophosphatidic acid (LPA). Thus, in these cells, LPA activation of PAK is mediated by G α i and is dependent on Src. In addition, activation of PAK by LPA requires generation of ROS.⁵²

Tissue factor activates the extrinsic pathway of coagulation, which leads to thrombin formation. Thrombin induces expression of tissue factor mRNA in vascular smooth muscle cells and, consequently, contributes to prolonged procoagulant activity and increased thrombogenicity at sites of vascular lesion. NAD(P)H oxidase participates in the redox-sensitive induction of tissue factor mRNA expression and the surface procoagulant activity activated by thrombin. This response is mediated by the NAD(P)H oxidasedependent activation of MAPK and by the PI3K/protein kinase B pathway. Given that active tissue factor promotes the formation of thrombin, NAD(P)H oxidase could play a crucial role in perpetuating thrombogenic effects in lesioned vessel walls.⁵³

Phagocytes produce O_2^- through the reduction of molecular oxygen by the NAD(P)H oxidase complex. This complex is formed from a membrane-bound flavocytochrome and 3 cytosolic proteins, one of which is a dimer of Rac1 p21 or Rac2 p21 and the Rho-guanine nucleotide dissociation inhibitor (RhoGDI). It has been proposed that, following dissociation of RhoGDI, Rac1 p21 bound to GDP is the physiologic activator of NAD(P)H-oxidase in macrophages, and that nucleotide exchange or the conversion of GTP are not necessarily involved.⁵⁴

The role of the Rho kinase p160-ROCK in the production of O_2^- by human polymorphonuclear leukocytes, and in the aggregation and adhesion of this cell type has been investigated under physiologic conditions using the selective p160-ROCK inhibitor Y-27632.⁵⁵ Here it was observed that production of O_2^- stimulated by phorbol ester was inhibited by Y-27632 in a concentration-dependent manner, indicating that p160-ROCK participated in this process and in the aggregation of human polymorphonuclear leukocytes.

Hydrogen peroxide, a reactive oxygen species, causes pulmonary edema, and increases hydrostatic pressure and vascular permeability. Increased permeability is accompanied by contraction and reorganization of the cytoskeleton in endothelial cells that leads to the formation of intercellular adhesions. The Rho family is also implicated in cell contraction. It has been demonstrated that an inhibitor of Rho kinase blocks the generation of this type of edema in rabbit lung, antagonizing the effects of H_2O_2 . This finding indicates that Rho is involved in H_2O_2 -induced pulmonary edema.⁵⁶

PLEIOTROPIC EFFECTS OF THE STATINS THROUGH THE RHO SIGNALING PATHWAY

Statins, inhibitors of HMG-CoA reductase, are potent inhibitors of cholesterol biosynthesis. A number of clinical studies have demonstrated beneficial effects of statins in the primary and secondary prevention of heart disease. However, their action seems to extend beyond what would be expected on the basis of hypolipidemic effects alone. Recent experimental and clinical evidence reveals that the pleiotropic effects of statins are independent of their actions on cholesterol and involve an improvement or restoration of endothelial function, increasing plaque stability and reducing oxidative stress and vascular inflammation. A number of the pleiotropic effects of the statins are mediated by their capacity to block the synthesis of important isoprenoid intermediates (farnesyl or geranylgeranyl) that serve as lipid anchors for various intracellular signaling molecules. In particular, the inhibition of small GTPases (Figure 4), whose membrane localization and function depend on the process of isoprenylation (specifically farnesylation or geranylgeranylation), would play an important role in mediating the direct cellular effects of the statins on the vessel wall.57

Despite all of the recent findings that have been discussed here, a number of questions remain to be answered in the coming years. It will be important to address whether similar patterns of cardiovascular activation of the Rho signaling pathway occur in different experimental models of arterial hypertension, to identify the main stimuli, and to assess how activation of the pathway is linked to the expression of genes that encode proteins with proinflammatory, profibrotic, and prothrombotic effects on the arteries. Furthermore, it will be of interest to determine whether there is any relationship between vascular activation of Rho signaling and local or systemic changes in vasoactive peptides, cell adhesion molecules, and/or prooxidant pathways in the development of arterial hypertension. Finally, it will be important to determine whether inhibition of distinct points in the Rho signaling pathway, once activated, affects the process of vascular remodeling in arterial hypertension or only blood pressure, given the many possible therapeutic targets beyond arterial hypertension (Table 2 and Figure 4).

In conclusion, the Rho/Rho kinase signal transduction pathway represents a newly recognized mechanism of vasoconstriction in arterial hypertension, pathologic cardiovascular remodeling, and a number of other cardiovascular and noncardiovascular conditions, and may represent a novel and promising therapeutic target.

REFERENCES

- Collins R, MacMahon S. Blood pressure, antihypertensive drug treatment and the risks of stroke and of coronary heart disease. Br Med Bull. 1994;50:272-98.
- Johns DG, Dorrance AM, Leite R, Weber DS, Webb RC. Novel signaling pathways contributing to vascular changes in hypertension. J Biomed Sci. 2000;7:431-43.
- Uehata M, Ishizaki T, Satoh H, Ono T, Kawahara T, Morishita T, et al. Calcium sensitization of smooth muscle mediated by a Rho-associated protein kinase in hypertension. Nature. 1997; 389:990-4.
- Kaibuchi K, Kuroda S, Amano M. Regulation of the cytoskeleton and cell adhesion by the Rho family GTP-ases in mammalian cells. Annu Rev Biochem. 1999;68:459-86.

TABLE 2. Diseases in Which Rho/Rho Kinase Could Represent a Therapeutic Target

Cardiovascular diseases Arterial hypertension Cerebral and coronary vasospasm Postangioplasty restenosis Pulmonary hypertension Erectile dysfunction Noncardiovascular diseases Bronchial asthma Premature birth Cancer (angiogenesis, cell proliferation, and motility and migration of tumor cells)

Taken from Somlyo et al.59



- Figure 4. Inhibition of the Rho kinase pathway and potential cardiovascular therapeutic effects. The addition of a farnesyl group to the small GTPases, necessary for their association with the plasma membrane and physiologic activity, can be blocked by statins and farnesyltransferase inhibitors (FTI). Statins inhibit HMG-CoA reductase, a key enzyme in the regulation of isoprenoid synthesis from acetyl-CoA, while FTIs directly inhibit far-nesyltransferase. Y-27632 and fasudil inhibit Rho kinase. All of these are potential therapeutic targets for the treatment of cardiovascular diseases.
- Mukai Y, Shimokawa H, Matoba T, Kandabashi T, Satoh S, Hiroki J, et al. Involvement of Rho-kinase in hypertensive vascular disease: a novel therapeutic target in hypertension. FA-SEB J. 2001;15:1062-4.
- Boguski MS, McCormick F. Proteins regulating Ras and its relatives. Nature. 1993;366:643-54.
- Clerk A, Sugden PH. Small guanine nucleotide-binding proteins and myocardial hypertrophy. Circ Res. 2000;86:1019-23.
- Aspenstro MP. Effectors for the Rho GTPases. Curr Opin Cell Biol. 1999;11:95-102.
- Vojtek AB, Der CJ. Increasing complexity of the Ras signaling pathway. J Biol Chem. 1998;273:19925-8.
- Maekawa M, Ishizaki T, Boku S, Watanabe N, Fujita A, Iwamatsu A, et al. Signaling from Rho to the actin cytoskeleton through protein kinases ROCK and LIM-kinase. Science. 1999;285:895-8.
- Carlier MF, Ressad F, Pantaloni D. Control of actin dynamics in cell motility: role of ADF/cofilin. J Biol Chem. 1999; 274:33827-30.
- Kuwahara K, Saito Y, Nakagawa O, Kishimoto I, Harada M, Ogawa E, et al. The effects of the selective ROCK inhibitor, Y27632, on ET-1-induced hypertrophic response in neonatal

rat cardiac myocytes: possible involvement of Rho/ROCK pathway in cardiac muscle cell hypertrophy. FEBS Lett. 1999;452:314-8.

- Zohn IM, Campbell SL, Khosravi-Far R, Rossman KL, Der CJ. Rho family proteins and Ras transformation: the RHOad less traveled gets congested. Oncogene. 1998;17:1415-38.
- Benard V, Bokoch GM, Diebold BA. Potential drug targets: small GTPases that regulate leukocyte function. Trends Pharmacol Sci. 1999;20:365-70.
- Galan JE. Interactions of salmonella with host cells: encounters of the closest kind. Proc Natl Acad Sci USA. 1998;95:14006-8.
- 16. Finkel T. Signal transduction by reactive oxygen species in non-phagocytic cells. J Leukoc Biol. 1999;65:337-40.
- Seasholtz TM, Majumdar M, Brown JH. Rho as a mediator of G protein coupled receptor signaling. Mol Pharmacol. 1999;55:949-56.
- Matrougui K, Tanko LB, Loufrani L, Gorny D, Levy BI, Tedgui A, et al. Involvement of Rho-kinase and the actin filament network in angiotensin II-induced contraction and extracellular signal-regulated kinase activity in intact rat mesenteric resistance arteries. Arterioscler Thromb Vasc Biol. 2001;21:1288-93.

- Chrissobolis S, Sobey CG. Evidence that Rho-kinase activity contributes to cerebral vascular tone in vivo and is enhanced during chronic hypertension. Circ Res. 2001;88:774-9.
- Kobayashi N, Nakano S, Mita S, Kobayashi T, Honda T, Tsubokou Y, et al. Involvement of Rho-kinase pathway for angiotensin II-induced plasminogen activator inhibitor-1 gene expression and cardiovascular remodeling in hypertensive rats. J Pharmacol Exp Ther. 2002;301:459-66.
- Kataoka C, Egashira K, Inoue S, Takemoto M, Ni W, Koyanagi M, et al. Important role of Rho-kinase in the pathogenesis of cardiovascular inflammation and remodeling induced by longterm blockade of nitric oxide synthesis in rats. Hypertension. 2002;39:245-50.
- Wesselman JP, de Mey JG. Angiotensin and cytoskeletal proteins: role in vascular remodeling. Curr Hypertens Rep. 2002;4:63-70.
- Aoki H, Izumo S, Sadoshima J. Angiotensin II activates RhoA in cardiac myocytes: a critical role of RhoA in angiotensin IIinduced premyofibril formation. Circ Res. 1998;82:666-76.
- Aikawa R, Komuro I, Yamazaki T, Zou Y, Kudoh S, Zhu W, et al. Rho family of small G proteins play critical roles in mechanical stress-induced hypertrophic responses in cardiac myocytes. Circ Res. 1999;84:458-66.
- Aikawa R, Komuro I, Nagai R, Yazaki Y. Rho plays an important role in angiotensin II-induced hypertrophic responses in cardiac myocytes. Mol Cell Biochem. 2000;212:177-82.
- 26. Yamakawa T, Tanaka S, Numaguchi K, Yamakawa Y, Motley ED, Ichihara S, et al. Involvement of Rho-kinase in angiotensin II-induced hypertrophy of rat vascular smooth muscle cells. Hypertension. 2000;35:313-8.
- Numaguchi K, Eguchi S, Yamakawa T, Motley ED, Inagami T. Mechanotransduction of rat aortic vascular smooth muscle cells requires RhoA and intact actin filaments. Circ Res. 1999;85:5-11.
- Funakoshi Y, Ichiki T, Shimokawa H, Egashira K, Takeda K, Kaibuchi K, et al. Rho-kinase mediates angiotensin II-induced monocyte chemoattractant protein-1 expression in rat vascular smooth muscle cells. Hypertension. 2001;38:100-4.
- Pan ZK, Ye RD, Christiansen SC, Jagels MA, Bokoch GM, Zuraw BL. Role of the Rho GTPase in bradykinin-stimulated nuclear factor-kappaB activation and IL-1beta gene expression in cultured human epithelial cells. J Immunol. 1998;160:3038-45.
- 30. Seasholtz TM, Zhang T, Morissette MR, Howes AL, Yang AH, Brown JH. Increased expression and activity of RhoA are associated with increased DNA synthesis and reduced p27Kip1 expression in the vasculature of hypertensive rats. Cir Res. 2001;89: 488-95.
- Chitaley K, Webb RC. Nitric oxide induces dilation of rat aorta via inhibition of rho-kinase signaling. Hypertension. 2002;39: 438-42.
- Sauzeau V, Rolli-Derkinderen M, Marionneau C, Loirand G, Pacaud P. RhoA expression is controlled by nitric oxide through cGMP-dependent protein kinase activation. J Biol Chem. 2003;278:9472-80.
- van Wetering S, van Buul JD, Quik S, Mul FP, Anthony EC, Klooster JP, et al. Reactive oxygen species mediate Rac-induced loss of cell-cell adhesion in primary human endothelial cells. J Cell Sci. 2002;115:1837-46.
- Hernández-Perera O, Pérez-Sala D, Soria E, Lamas S. Involvement of Rho GTPases in the transcriptional inhibition of preproendothelin-1 gene expression by simvastatin in vascular endothelial cells. Circ Res. 2000;87:616-22.
- 35. Adamson RH, Curry FE, Adamson G, Liu B, Jiang Y, Aktories K, et al. Rho and rho kinase modulation of barrier properties: cultured endothelial cells and intact microvessels of rats and mice. J Physiol. 2002;539:295-308.
- Verin AD, Birukova A, Wang P, Liu F, Becker P, Birukov K, et al. Microtubule disassembly increases endothelial cell barrier dysfunction: role of MLC phosphorylation. Am J Physiol Lung Cell Mol Physiol. 2001;281:L565-74.

- 37. Soga N, Namba N, McAllister S, Cornelius L, Teitelbaum SL, Dowdy SF, et al. Rho family GTPases regulate VEGFstimulated endothelial cell motility. Exp Cell Res. 2001;269: 73-87.
- Ashida N, Arai H, Yamasaki M, Kita T. Distinct signaling pathways for MCP-1-dependent integrin activation and chemotaxis. J Biol Chem. 2001;276:16555-60.
- Takeda K, Ichiki T, Tokunou T, Iino N, Fujii S, Kitabatake A, et al. Critical role of Rho-kinase and MEK/ERK pathways for angiotensin II-induced plasminogen activator inhibitor type-1 gene expression. Arterioscler Thromb Vasc Biol. 2001;21:868-73.
- 40. Choi SE, Choi EY, Kim PH, Kim JH. Involvement of protein kinase C and rho GTPase in the nuclear signalling pathway by transforming growth factor-beta1 in rat-2 fibroblast cells. Cell Signal. 1999;11:71-6.
- Laudanna C, Campbell JJ, Butcher EC. Role of Rho in chemoattractant-activated leukocyte adhesion through integrins. Science. 1996;271:981-3.
- Ronald JA, Ionescu CV, Rogers KA, Sandig M. Differential regulation of transendothelial migration of THP-1 cells by ICAM-1/LFA-1 and VCAM-1/VLA-4. J Leukoc Biol. 2001; 70:601-19.
- 43. Takeuchi S, Kawashima S, Rikitake Y, Ueyama T, Inoue N, Hirata K, et al. Cerivastatin suppresses lipopolysaccharide-induced ICAM-1 expression through inhibition of Rho GTPase in BAEC. Biochem Biophys Res Commun. 2000;269:97-102.
- Wojciak-Stothard B, Williams L, Ridley AJ. Monocyte adhesion and spreading on human endothelial cells is dependent on Rho-regulated receptor clustering. J Cell Biol. 1999;145:1293-307.
- 45. Etienne S, Adamson P, Greenwood J, Strosberg AD, Cazaubon S, Couraud PO. ICAM-1 signaling pathways associated with Rho activation in microvascular brain endothelial cells. J Immunol. 1998;161:5755-61.
- 46. Adamson P, Etienne S, Couraud PO, Calder V, Greenwood J. Lymphocyte migration through brain endothelial cell monolayers involves signaling through endothelial ICAM-1 via a rhodependent pathway. J Immunol. 1999;162:2964-73.
- 47. Walters CE, Pryce G, Hankey DJ, Sebti SM, Hamilton AD, Baker D, et al. Inhibition of Rho GTPases with protein prenyltransferase inhibitors prevents leukocyte recruitment to the central nervous system and attenuates clinical signs of disease in an animal model of multiple sclerosis. J Immunol. 2002;168:4087-94.
- del Pozo MA, Vicente-Manzanares M, Tejedor R, Serrador JM, Sánchez-Madrid F. Rho GTPases control migration and polarization of adhesion molecules and cytoskeletal ERM components in T lymphocytes. Eur J Immunol. 1999;29:3609-20.
- Kaneider NC, Reinisch CM, Dunzendorfer S, Romisch J, Wiederman CJ. Syndecan-4 mediates antithrombin-induced chemotaxis of human peripheral blood lymphocytes and monocytes. J Cell Sci. 2002;115:227-36.
- 50. Eto M, Barandier C, Rathgeb L, Kozai T, Joch H, Yang Z, et al. Thrombin suppresses endothelial nitric oxide synthase and upregulates endothelin-converting enzyme-1 expression by distinct pathways: role of Rho/ROCK and mitogen-activated protein kinase. Circ Res. 2001;89:583-90.
- Mizuno T, Kaibuchi K, Ando S, Musha T, Hiraoka K, Takaishi K, et al. Regulation of the superoxide-generating NADPH oxidase by a small GTP-binding protein and its stimulatory and inhibitory GDP/GTP exchange proteins. J Biol Chem. 1992; 267:10215-8.
- Schmitz U, Thommes K, Beier I, Vetter H. Lysophosphatidic acid stimulates p21-activated kinase in vascular smooth muscle cells. Biochem Biophys Res Commun. 2002;291:687-91.
- Herkert O, Diebold I, Brandes RP, Hess J, Busse R, Gorlach A. NADPH oxidase mediates tissue factor-dependent surface procoagulant activity by thrombin in human vascular smooth muscle cells. Circulation. 2002;105:2030-6.

- 54. Bromberg Y, Shani E, Joseph G, Gorzalczany Y, Sperling O, Pick E. The GDP-bound form of the small G protein Rac1 p21 is a potent activator of the superoxide-forming NADPH oxidase of macrophages. J Biol Chem. 1994;269:7055-8.
- 55. Kawaguchi A, Ohmori M, Harada K, Tsuruoka S, Sugimoto K, Fujimura A. The effect of a Rho kinase inhibitor Y-27632 on superoxide production, aggregation and adhesion in human polymorphonuclear leukocytes. Eur J Pharmacol. 2000;403: 203-8.
- Chiba Y, Ishii Y, Kitamura S, Sugiyama Y. Activation of rho is involved in the mechanism of hydrogen-peroxide-induced lung

edema in isolated perfused rabbit lung. Microvasc Res. 2001; 62:164-71.

- Takemoto M, Liao JK. Pleiotropic effects of 3-hydroxy-3methylglutaryl coenzyme A reductase inhibitors. Arterioscler Thromb Vasc Biol. 2001;21:1712-9.
- Lee DL, Webb RC, Jin L. Hypertension and RhoA/Rho-Kinase signaling in the vasculature. Highlights from the recent literature. Hypertension. 2004;44:796-9.
- 59. Somlyo AP, Somlyo AV. Ca+2 sensitivity of smooth muscle and nonmuscle myosin II: modulated by G proteins, kinases, and myosin phosphatase. Physiol Rev. 2003;83:1325-58.

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